256. Synthesis of Cannabinoid Model Compounds

Part 31)

(6aR, 10aR)-N-Ethyl-∆⁸-tetrahydrocannabinol-18-amide, (6aR, 10aR, 17RS)-N-Ethyl-17-methyl-∆⁸-tetrahydrocannabinol-18-amide and (6aR, 10aR)-17,18-Didehydro-∆⁸-tetrahydrocannabinol

by Burkhard Schmidt, Ingo Franke, Franz-Josef Witteler and Michael Binder*

Institut für Physiologische Chemie der Ruhr-Universität Bochum, Postfach 102148, D-4630 Bochum 1

Dedicated to Prof. Dr. Ch. Tamm on the occasion of his 60th birthday

(1.IX.83)

Summary

The novel cannabinoids (6aR, 10aR)-N-ethyl- Δ^8 -tetrahydrocannabinol-18-amide (15) and (6aR, 10aR, 17RS)-N-ethyl-17-methyl- Δ^8 -tetrahydrocannabinol-18-amide (16), designed as cannabinoid affinity ligands, were synthesized from the corresponding acids 11 and 12 via the N-hydroxysuccinimide esters. Amide 16 was tested in the rat and was generalized to Δ^9 -tetrahydrocannabinol, being 5 times less potent than the training drug. An improved synthesis of (6aR, 10aR)-17,18-didehydro- Δ^8 -tetrahydrocannabinol (23) is reported. As model reaction for the preparation of a tritiated Δ^8 -tetrahydrocannabinol, compound 23 was selectively deuterated at C(17) and C(18) in benzene/Et₃N using [(C₆H₅)₃P]₃RuCl₂ as catalyst.

Recently, we have reported the synthesis of (6aR, 10aR)- Δ^8 -tetrahydrocannabinol-18oic acid²) (= Δ^8 -THC-18-oic acid; 11) and (17 RS)-17-methyl- Δ^8 -THC-18-oic acid (12) [1]. The latter compound was designed as an affinity ligand to isolate Δ^9 (or Δ^8)-THCbinding macromolecules (the 'THC receptor' [2]) from neuronal membrane fractions. Acid 12 retains the full structure of Δ^9 -THC and was to be coupled to BrCN-activated Sepharose via an ethylamino spacer. The closest model to the moiety attached to the Sepharose is the N-ethyl amide of acid 12. To evaluate efficient techniques for coupling the acid 12 to the aminoethyl-Sepharose and to study whether the amide retains psychotropic activity, *i.e.* binds to the THC receptor, the N-ethyl amides 15 and 16 of acids 11 and 12, respectively, were synthesized.

¹) Part 2: see [1].

²) This numbering of the cannabinoid skeleton is based on a suggestion by *Schoenfeld* [3]. It is semisystematic. The numbering of the ring system follows the IUPAC rule but the substituents are numbered consecutively.

Characterization of Δ^9 -THC-binding sites requires radiolabelled ligands of high specific activity and 17,18-didehydro- Δ^8 -THC (23) was chosen as precursor for the synthesis of $[17,18^{-3}H_2]$ - Δ^8 -THC (24). This paper describes also the synthesis of 23 and 24.



1. Syntheses of Amides 15 and 16 (Scheme 1). – Basically, the acids 11 and 12 were obtained as described in [1] $(1 + 2 \rightarrow 4 \rightarrow 6(+8) \rightarrow 9 \rightarrow 11; 1 + 3 \rightarrow 5 \rightarrow 7(+8) \rightarrow 10 \rightarrow 12)$. However, the yield of Wittig adduct 5 was increased to 37% by substituting the phosphonium bromide [1] by the corresponding iodide 3. Since both amides and acids were intended as model compounds for the affinity gels we restricted ourselves in transforming 11 and 12 into 15 and 16, respectively, to conditions that would allow the coupling of the acids to the activated Sepharose as well. The THC molecule is very reactive, especially involvement of the free phenolic OH group has to be avoided. The use of 1-ethyl-3-[3'-(dimethylamino)propyl]carbodiimide for the amide formation proved highly unsatisfactory; both amides 15 and 16 were obtained in yields of 7%. Following the method of Cook [4], 15 and 16 were obtained in 70% yield by reacting 11 and 12, respectively, with N-hydroxysuccinimide in anhydrous MeCN in the presence of dicyclohexylcarbodiimide (DCC) and treating the intermediate esters 13 and 14, respectively, with EtNH₂ in aqueous dioxane.



Fig. 1. Fragments (m/z) in the MS of the Amides 15 and 16. Values of 16 in italics, relative intensities in parentheses.

Amide 16 was tested for psychotropic activity in the rat [5]. In a drug-discrimination paradigm the compound was generalized to Δ^9 -THC at 5 times the concentration of the training drug. This proves that the model compound retains affinity to the postulated THC receptor and justifies our choice of acid 12 as affinity ligand. Metabolic transformation, pharmacokinetics and triglyceride/phospholipid partitioning of amide 16 are investigated at present and will be reported elsewhere.

The structures of the amides 15 and 16 were proven by their mass spectra and ¹H-NMR spectra. The MS are given in *Fig. 1*. In addition to the known fragmentations of the isoprenoid part of the molecule [6] [7], prominent fragment ions arise from cleavage of the substituted pentyl side chain. Decoupling allowed to localize all signals of the 250-MHz NMR spectrum of 16 (*Fig. 2*).

Since the ¹H-NMR spectra of tetrahydrocannabinols have been analyzed in detail [8] only signals relevant to the substituted side chain will be discussed here. The CH₃CH₂NHCO-function gives rise to a dq (CH₃CH₂) centered at 3.28 and 3.25, a *t* at 5.58 (NH), and a *t* at 1.1 ppm (CH₃CH₂). The protons at C(15) were located as a quint ($J_{14,15} = J_{15,16} = 7$ Hz) at 1.54 ppm by irradiation at 2.42 ppm, the position of the signals of H–C(14). Irradiation at 1.54 ppm revealed the location of the 2 H–C(16) at 1.41 ppm. The CH₃–C(17) leads to a dat 1.1 ppm. By decoupling at this position, H–C(17) was located at 2.14 ppm.



Fig. 2. 250-MHz ¹H-NMR Spectrum of (6aR, 10aR, 17RS)-N-Ethyl-17-methyl- Δ^8 -tetrahydrocannabinol-18-amide (16) in $CDCl_3$

2. Synthesis of 17,18-Didehydro- Δ^8 -THC (23) and $[17,18-^2H_2]-\Delta^8$ -THC (24) (Scheme 2). - Essentially following the procedure of *Pitt et al.* [9], several changes were introduced. The 3,5-dimethoxybenzaldehyde (17) was reacted with the triphenylphosphonium bromide 18 in Et₂O in the presence of BuLi to give a *cis/trans*-mixture of the *Wittig* adduct 19 (62%). This reaction did not work at the concentrations given in [9] but proceeded smoothly at 10fold dilution. Catalytic hydrogenation of 19 (Pd/C, EtOH) gave 20 in 95% yield. Using BBr₃ in benzene, 20 was transformed into 5-(5-bromopentyl)resorcinol (21; 80%). Condensation of 21 with (+)-trans-p-mentha-2,8-dien-1-ol (8) yielded (6aR, 10aR)-18-bromo- Δ^8 -THC (22; 38%). In contrast to Pitt et al. [9] we succeeded in isolating 17,18-didehydro- Δ^8 -THC (23) in 14% yield from the mixture of products resulting from the dehydrohalogenation of 22 with potassium 3-ethylpentan-3-olate in benzene/toluene. The synthesis of 17,18-didehydro- Δ^9 -THC described by *Pitt et al.* [9] by adding HCl to the 8,9-double bond of 23 followed by dehydrohalogenation could not be reproduced in our laboratory. Finally, 17,18-didehydro- Δ^8 -THC (23) was deuterated selectively in benzene/Et₃N in the presence of $[(C_6H_5)_3P]_3RuCl_2$ to give $[17,18^{-2}H_2]-4^8$ -THC (24) in 49% yield. The essential advantage of this synthesis becomes evident: by introducing the radioactive isotope ${}^{3}H$ in the last reaction, extensive handling of radiolabelled compounds can be avoided. The structures of all synthetic intermediates were established by MS and ¹H-NMR spectroscopy.

The MS of 17,18-didehydro- \varDelta^8 -THC (**23**; M^{\ddagger} at m/z 312) closely resembles the one of \varDelta^8 -THC except for the base peak which arises from the cleavage between C(14) and C(15) including a *McLafferty* rearrangement (m/z 258; M^{\ddagger} -C₄H₆). The MS of the deuterated derivative **24** revealed 15.6% non-, 27.3% mono-, 32.4% bi-, 15.3% tri-, 6.3%



tetra- and 2.1% pentadeuteration, giving a total of 1.73 ²H per molecule. The ¹H-NMR spectrum (*Fig. 3*) of **23** reveals 3 additional signals of olefinic protons between 4.8 and 6.0 ppm arising from H–C(17) and H–C(18) while the usual t of H–C(18) of Δ^8 -THC at 0.9 ppm is lacking. Broad signals at 0.85 and 1.29 ppm in the 250-MHz ²H-NMR spectrum of **24** indicate the presence of deuterium atoms mostly at C(18) and C(16/17).

The authors are indebted to Dr. W. Dietrich, Dept. of NMR spectroscopy, Ruhr-University, and Dr. D. Müller, Dept. of mass spectroscopy, Ruhr-University, for recording the analytical data. The work reported here was supported by a grant from the Deutsche Forschungsgemeinschaft.



Fig. 3. 80-MHz ¹H-NMR Spectrum of 17, 18-Didehydro- Δ^{8} -THC (23) in CDCl₃

Experimental Part

General Remarks. See [1].

1. (6aR, 10aR)-N-Ethyl- Δ^8 -tetrahydrocannabinol-18-amide (= N-Ethyl-5-[(6aR, 10aR)-1-hydroxy-6,6,9-trimethyl-6a,7,10,10a-tetrahydro-6 H-dibenzo[b,d]pyran-3-yl]pentanamide; **15**) and (6aR, 10aR, 17RS)-N-Ethyl-17methyl- Δ^8 -tetrahydrocannabinol-18-amide (= (2RS)-N-Ethyl-5-[(6aR, 10aR)-1-hydroxy-6,6,9-trimethyl-6a,7,10,10atetrahydro-6 H-dibenzo[b,d]pyran-3-yl]-2-methylpentanamide; **16**). Only procedures that are not identical to those already published [1] are given in detail.

1.1. (6 aR,10 aR)- A^8 -Tetrahydrocannabinol-18-oic Acid (= 5-[(6 aR,10 aR)-1-Hydroxy-6,6,9-trimethyl-6 a,7, 10,10 a-tetrahydro-6 H-dibenzo[b,d]pyran-3-yl]pentanoic Acid; 11). See [1].

1.2. (6 aR, 10 aR, 17RS)-17-Methyl- A^8 -tetrahydrocannabinol-18-oic Acid (= (2RS)-5-[(6 aR, 10 aR)-1-Hydroxy-6,6,9-trimethyl-6 a,7,10,10a-tetrahydro-6H-dibenzo[b,d]pyran-3-yl]-2-methylpentanoic Acid; **12**). – 1.2.1. Methyl (2RS)-4-Iodo-2-methylbutanoate. A solution of 2.15 g (11 mmol) of methyl (2RS)-4-bromo-2-methylbutanoate [1] in 7.5 ml of dry acetone was added dropwise to 2.47 g (16.5 mmol) of NaI in 30 ml of dry acetone, and the mixture was heated under reflux for 90 min. The solvent was removed under N₂ and the residue partitioned between Et₂O/H₂O. The org. phase yielded 2.62 g (98%) of pure methyl 4-iodo-2-methylbutanoate. ¹H-NMR: 1.17 (d, J = 7, 3H, CH₃-C(2)); 1.7 (m, 2H, 2H-C(3)); 2.4–2.8 (m, 1H, H-C(2)); 3.20 (t, J = 6, 2H, 2H-C(4)); 3.61 (s, 3H, CH₃O). MS: 211 (13, M⁺-OCH₃), 183 (9, M⁺-COOCH₃), 155 (6, M⁺-CH(CH₃)COOCH₃), 115 (75, M⁺-I), 88 (16, CH₃CH=C(OH)OCH₃⁺), 59 (100, COOCH₃⁺).

1.2.2. (3-(Methoxycarbonyl)butyl)triphenylphosphonium Iodide (3). A solution of 2.62g (10.8 mmol) of methyl (2RS)-4-iodo-2-methylbutanoate and 2.27g (8.65 mmol) of Ph₃P in 20 ml of dry benzene was heated under reflux for 65 h. The resinous 3 was allowed to separate, washed 5 × with benzene (30 ml) and dried at 0.01 Torr: 4.27g (98%). ¹H-NMR ((D₆)DMSO): 1.16 (d, J = 7, 3H, CH₃-C(2)); 1.5-2.1 (m, 2H, 2H-C(3)); 3.1-3.4 (m, 2H, 2H-C(4)); 3.63 (s, 3H, CH₃O); 3.4-3.8 (m, 1H, H-C(2)); 7.8 (m, 15H).

1.2.3. Methyl (2RS,4EZ)-5-[3',5'-bis(benzyloxy)phenyl]-2-methyl-4-pentenoate (5). K (350 mg, 9 mmol) was reacted with dry 2-methyl-2-butanol (15 ml). The excess alcohol was destilled off an the alcoholate was dissolved in

15 ml of hexamethylphosphoric triamide. At 0°, a solution of 4.27 g (8.5 mmol) of 3 in 20 ml of dry DMF was added with simultaneous formation of the orange ylide. After 15 min, a solution of 2.0 g (6.3 mmol) of 3,5-bis(benzyloxy)benzaldehyde (1) in 20 ml of benzene was introduced. The mixture was stirred for 2 h at 23° and partitioned between Et_2O/H_2O . The crude product obtained from the Et_2O -phase was dissolved in 50 ml of CCl₄, and the Ph₃PO was precipitated by addition of 300 ml of petroleum ether. Chromatography of the product on 200 g of silica gel (fract. size 200 ml, toluene/petroleum ether 2:1) yielded 1.316 g (37% calc. on 3) of pure 5 (fract. 18–27). Analytical data: see [1].

1.2.4. Transformations from 5 to 7, 7 to 10 and 10 to 12. See [1].

1.3. Amide **16** from **12**. A solution of 589 mg (2.86 mmol) of dicyclohexylcarbodiimide (DCC) in 15 ml of abs. MeCN was added dropwise at 0° to 1.025 g (2.86 mmol) of **12** and 329 mg (2.86 mmol) of *N*-hydroxysuccinimide in 13 ml of abs. MeCN. Precipitation of dicylohexylurea started immediately. The mixture was stirred at 23° for 24 h, the urea removed by filtration and solvent removed *in vacuo*. The crude ester **14** was dissolved in 16 ml of dioxane and added at 23° to a solution of 420 mg of NaHCO₃ and 255 µl of EtNH₂ (70% in H₂O; 3.15 mmol) in 35 ml of H₂O/dioxane 3 : 2. Stirring was continued for 90 min, the solvent removed *in vacuo*, and the residue partitioned between Et₂O/H₂O. The crude product resulting from the org. phase was adsorbed to 10g of silica gel and chromatographed on 90 g of silica gel (fract. size 100 ml; fract. 1–20 Et₂O/petroleum ether 1 : 1, fract. 20–40 Et₂O). Fract. 21–27 yielded 887 mg (77%) of pure **16**. ¹H-NMR (*cf. Fig. 2*): 1.068 (*s*, 3H, 3H–C(13)); 1.098 (*t*, *J* = 7, 3H, NCH₂CH₃); 1.36 (*s*, 3H, 3H–C(6a), H_a–C(7), H_b–C(10)); 1.54 (*quint.*, *J* = 7, 2H, 2H–C(15)); 1.68 (*s*, 3H, 3H–C(11)); 1.78 (*m*, 3H, H–C(6a), H_a–C(7), H_b–C(10)); 2.74 (*m*, 2H, H_b–C(7), H_c–C(10)); 5.58 (*t*, *J* = 7, NH); 6.15, 6.18 (2*d*, *J* = 2 each, 2H, H–C(2), H–C(2)). MS: see *Fig. J*, values in italics.

1.4. Amide 15 from 11. As described in 1.3, 356 mg (0.98 mmol) of 11 were reacted with 110 mg (0.95 mmol) of N-hydroxysuccinimide and 196 mg (0.95 mmol) of DCC. The crude ester 13 was treated with 120 mg of NaHCO₃ and 80 μ l of 70% EtNH₂. Chromatography on silica gel yielded 308 mg (80%) of pure 15. ¹H-NMR: same signals as 16 except for CH₃-C(17), H-C(17) and H-C(16). MS: see Fig. 1.

2. (6aR, 10aR) - 17, 18-Didehydro- Δ^8 -tetrahydrocannabinol (= (6aR, 10aR) - 6, 6, 9-Trimethyl-3-(4-pentenyl)-6a, 7, 10, 10a-tetrahydro-6H-dibenzo[b,d]pyran-1-ol; **23**) and $(6aR, 10aR) - [17, 18 - 2H_2] - \Delta^8$ -Tetrahydrocannabinol (= (6aR, 10aR) - 6, 6, 9-Trimethyl-3- $([4, 5 - 2H_2] - 10, 10a$) and $(6aR, 10aR) - [17, 18 - 2H_2] - \Delta^8$ -Tetrahydrocannabinol (= (6aR, 10aR) - 6, 6, 9-Trimethyl-3- $([4, 5 - 2H_2] - 10, 10a$) and (6aR, 10aR) - 6H-dibenzo[b,d]pyran-1-ol; **24**). 2.1. 1 - (3', 5' - Dimethoxyphenyl) - 5-phenoxy-1-pentene (**19**). A solution of 2.6 ml ($\approx 4.2 \text{ mmol}$) of BuLi in hexane was added dropwise at 0° to a suspension of 2.0 g(4.1 mmol) of (4-phenoxybutyl)triphenylphosphonium bromide (**18**) in 50ml of dry Et₂O. After stirring the red mixture for 30min at 0°, a solution of 0.70g (4.2 mmol) of 3,5-dimethoxybenzaldehyde (**17**) in 20ml of Et₂O was slowly introduced. A pale yellow precipitate (Ph₃PO) formed, the mixture was allowed to warm up to 23°, and, after addition of 30ml of Et₂O, was heated under reflux for 2h. The solvent was removed *in vacuo* and the crude product partitioned between petroleum ether/H₂O. The product contained in the org. phase was chromatographed on silica gel (200g; fract. size 200 ml; fract. 1-10 petroleum ether/Et₂O98 : 2, fract. 11-20 petroleum ether/Et₂O95 : 5). Fract. 12 and 13 gave 230 mg of pure (Z)-19, fract. 14-19 530 mg of (E/Z)-19 (together 62% yield). ¹H-NMR ((Z)-19): 1.95 (quint., 2H, 2H-C(4)); 2.5 (q, 2H, 2H-C(3)); 3.8 (s, 6H, 2CH₃O); 4.0 (t, J = 7, 2H, 2H-C(5)); 5.7 (dt, J = 11.4, 7, 1H, H-C(2)); 6.2-7.4 (div. *m*, 9H). MS: 298 (100, M⁺), 205(83, M⁺-OC₆H₃). 191 (59, M⁺-CH₂OC₆H₅), 178 (76, M⁺ - H₂C = CHOC₆H₅); 151 (36, M⁺ - H₂C = CHOC₆H₅). 191 (59, M⁺ - CH₂OC₆H₅), 178 (76, M⁺ - H₂C = CHOC₆H₅); 151 (36, M⁺ - H₂C = CHOC₆H₅).

2.2. I-(3',5'-Dimethoxyphenyl)-5-phenoxypentane (20). Compound 19 (1.732 g, 5.81 mmol) was hydrogenated at 1.5 atm/23° over 10% Pd/C (200 mg) in 60 ml of EtOH for 1 h. The catalyst was removed by filtration and the solvent evaporated *in vacuo*: 1.656 g (95%) of pure 20. ¹H-NMR: 1.5-2.0 (*m*, 6H, 2H-C(2), 2H-C(3), 2H-C(4)); 2.62 (*t*, J = 7, 2H, 2H-C(1)); 3.82 (*s*, 6H, 2CH₃O); 3.97 (*t*, J = 7, 2H, 2H-C(5)); 6.4 (*m*, 3H, H-C(2'), H-C(4'), H-C(6')); 6.8-7.4 (*m*, 5H, OC₆H₅). MS: 300 (34, M^{\pm}); 207, (22, M^{\pm} -OC₆H₅); 152 (100, M^{\pm} - H₂CC=CHCH₂OC₆H₅); 151 (62, M^{\pm} - H₂CCH₂CH₂OC₆H₅).

2.3. 5-(5'-Bromopentyl) resorcinol (21). BBr₃ (0.83 ml; 8.7 mmol) in 30 ml of dry benzene was added dropwise at 10° to a stirred solution of 2.476 g (8.3 mmol) of 20 in 100 ml of dry benzene. Stirring was continued at 23° for 48 h with successive addition of BBr₃ (0.63 ml (16 h), 0.42 ml (18 h) and 0.2 ml (23 h)). The mixture was poured on 100 ml of ice H₂O and the org. phase was washed twice with 100 ml of 5% NaHSO₃. The solvent was removed *in vacuo* and the residue taken up in 10 ml of acetone and 100 ml of H₂O. Extraction with benzene yielded crude 21 that was chromatographed on silica gel (200 g; fract. size 200 ml; fract. 1–10 petroleum ether/CH₂Cl₂ 1:1, fract. 11–20

CH₂Cl₂, fract. 21-30 CH₂Cl₂/Et₂O 9:1). Fract. 21-28 gave 1.738 g (80.8%) of pure **21**. ¹H-NMR: 1.2-2.05 (*m*, 6H, 2H-C(2'), 2H-C(3'), 2H-C(4')); 2.53 (*t*, J = 7, 2H, 2H-C(1')); 3.42 (*t*, J = 6, 2H, 2H-C(5')); 5.18 (br. s, 2H, 2OH); 6.25 (s, 3H, H-C(2), H-C(4); H-C(6)). MS: 258 and 260 (7 and 7', resp., M^+), 179 (55, $M^+ - Br$, 124 (85, $M^+ - Br - C_4H_7$), 123 (100, $M^+ - Br - C_4H_8$).

2.4. (6aR, 10aR)-18-Bromo- Δ^8 -tetrahydrocannabinol (= (6aR, 10aR)-3-(5-Bromopentyl)-6,6,9-trimethyl-6a,7,10,10a-tetrahydro-6H-dibenzo[b,d]pyran-1-ol; **22**). A mixture of 1.39 g (5.37 mmol) of **21**, 1.06 g **8** (Firmenich, New York) and 200 mg of p-TsOH \cdot H₂O in 75 ml of dry benzene was heated under reflux for 4 h. The solvent was removed *in vacuo*. The residue was dissolved in 100 ml of Et₂O, washed with 10% NaHCO₃ and H₂O (3 ×). The crude product resulting from the Et₂O phase was chromatographed on silica gel (200 g; fract. size 200 ml; fract. 1–20 petroleum ether/CH₂Cl₂ 9:1, 21–30 8:2, 31–40 6:4) to give 800 mg (38.0%) of pure **22** (fract. 31–37). ¹H-NMR: 1.2 (s, 3H, 3H–C(13)); 1.4 (s, 3H, 3H–C(12)); 1.73 (br. s, 3H, 3H–C(11)); 2.48 (t, J = 6, 2H, 2H–C(14)); 2.7 (m, 1H, H–C(10a)); 3.22 (dd, J = 16, 4, 1H, H_a–C(10)); 3.42 (t, J = 7, 2H, 2H–C(18)); 4.73 (s, 1H, OH); 5.45 (br. s, 1H, H–C(8)); 6.1 (d, J = 2, 1H, H–C(2)); 6.28 (d, J = 2, 1H, H–C(4)). MS: 394, 392 (25, M⁺); 379, 377 (6, M⁺ – CH₃); 351, 349 (24, M⁺ – C₃H₇); 326, 324 (6, M⁺ – 68); 313 (100, M⁺ – Br); 311, 309 (49, M⁺ – 68 – 15); 258 (22, M⁺ – C₄H₇Br); 257 (8, M⁺ – C₄H₈Br).

2.5. 17,18-Didehydro Compound 23. A 0.15 M solution (52 ml) of potassium 3-ethyl-3-pentanolate in xylene was added under stirring to 860 mg (2.2 mmol) of 22 in 20 ml of dry benzene. The mixture was heated for 19 h at 70°, cooled, and saturated with CO₂ for 30 min. After addition of 30 ml of benzene, the mixture was extracted with H₂O (3 × 100 ml), the solvent evaporated *in vacuo*, and the residue was purified by column chromatography on silica gel (100 g; fract. size 100 ml; petroleum ether/CH₂Cl₂ 3:7). Fract. 3–6 gave 285 mg of crude 23 that was further purified on silica gel (35 g; fract. size 35 ml; fract. 1–20 petroleum ether/toluene 9:1, 21–30 petroleum ether/toluene 3:1, 31–40 petroleum ether/toluene/CH₂Cl₂ 5:1:4). Fract. 34–36 yielded 228 mg of 23, still containing impurities. Final purification was achieved on silica gel/AgNO₃ 9:1 (35 g; fract. size 35 ml; fract. 1–10 petroleum ether/Et₂O 98:2, 11–20 95:5, 21–30 90:10, 31–40 80:20, 41–60 60:40). Fract. 45–49 gave 92 mg (14%) of pure 23. ¹H-NMR: see *Fig. 3*. MS: 312 (83, M^{\pm}), 297 (7, M^{\pm} – 15), 269 (24, M^{\pm} – 43), 258 (100, M^{\pm} – C₄H₆), 243 (8, M^{\pm} – C₄H₆ – CH₃), 229 (73, M^{\pm} – 68–15).

2.6. Dideuterio Compound 24. A solution of 30 mg of 23, 8.5 mg of $[(C_6H_5)_3P]_3RuCl_2$ (*Fluka*) and 1.5 µl of Et₃N in 1.5 ml of benzene was treated with 2H_2 at 3 atm for 24 h at 23° (no starting material left). The catalyst was removed by chromatography on aluminium oxide (neutral, act. III; 7 g; fract. size 7 ml; petroleum ether/CH₂Cl₂ 3:1). Fract. 49–82 yielded 24 (16 mg) that was rechromatographed on *Florisil* (5 g; fract. size 5 ml; toluene) to give 15 mg (49%) of pure 24 (fract. 2–12). ¹H-NMR: 0.9 (m, 2H, 2H–C(18)); 1.12 (s, 3H, 3H–C(13)); 1.38 (s, 3H, 3H–C(12)); 1.72 (br. s, 3H, 3H–C(11)); 2.48 (t, J = 8, 2H, 2H–C(14)); 2.75 (m, 1H, H–C(10a)); 3.22 (dd, J = 17, 4, 1H, H_a–C(10)); 5.43 (br. s, 1H, H–C(8)); 6.11 (d, J = 2, 1H, H–C(2)); 6.28 (d, J = 2, 1H, H–C(4)). MS: 316 (M^+).

REFERENCES

- [1] I. Franke & M. Binder, Helv. Chim. Acta 63, 2508 (1980).
- [2] M. Binder & I. Franke, 'Neuroreceptors', ed. F. Hucho, Walter de Gruyter, Berlin & New York, 1982, p. 151.
- [3] R. Schoenfeld, J. Chem. Inf. Comput. Sci. 20, 65 (1980).
- [4] C.E. Cook, 'Cannabinoid Analysis in Physiological Fluids', ACS Symposium Series 98, ed. J.A. Vinson, Am. Chem. Soc., Washington, 1979, p. 137.
- [5] M. Binder & S. Koch, unpublished results.
- [6] U. Claussen, H.-W. Fehlhaber & F. Korte, Tetrahedron 22, 3535 (1966).
- [7] H. Budzikiewicz, R.T. Alpin, D.A. Lightner, C. Djerassi, R. Mechoulam & Y. Gaoni, Tetrahedron 21, 1881 (1965).
- [8] R.A. Archer, D.B. Boyd, P.V. Demarco, I.J. Thyminsky & N.L. Allinger, J. Am. Chem. Soc. 92, 5200 (1970).
- [9] C.G. Pitt, D.T. Hobbs, H. Schran, C.E. Twine & D.L. Williams, J. Labelled Compd. 11, 551 (1975).